

## Preliminary communication

### Stepwise degradation of moenomycin A

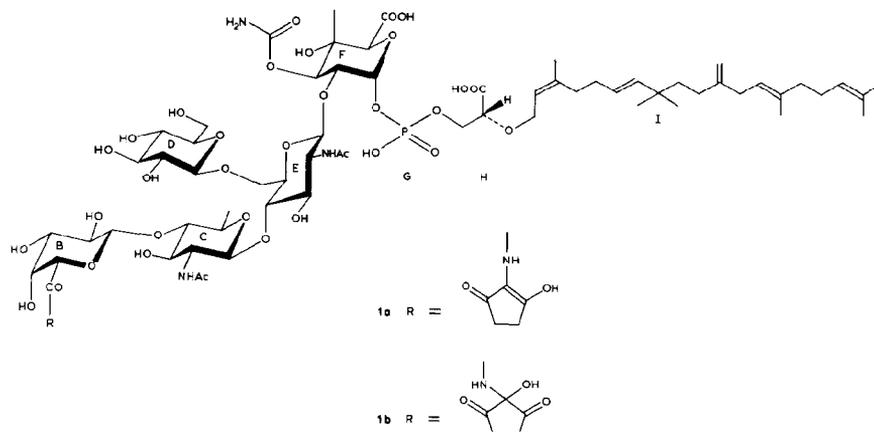
PETER WELZEL\*, FRANZ KUNISCH, FRITHJOF KRUGGEL, HERMANN STEIN, ARANKA PONTY, and HELMUT DUDDECK

*Abteilung für Chemie der Ruhr-Universität, Postfach 102148, D-4630 Bochum (West Germany)*

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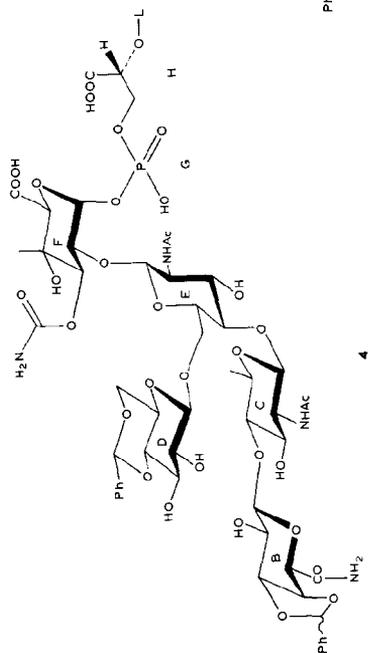
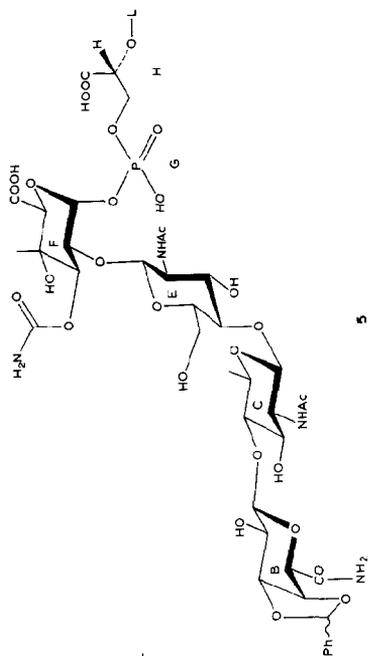
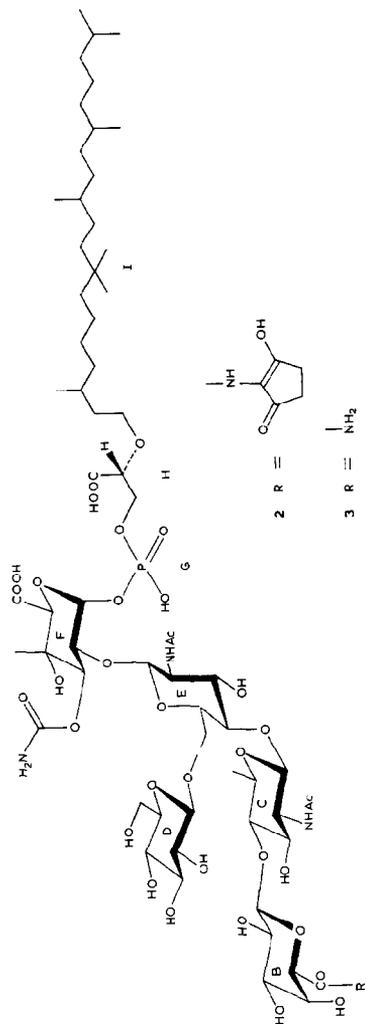
The antibiotic moenomycin A (**1a**) is the main constituent of the commercial product flavomycin<sup>®</sup> which is employed in animal nutrition<sup>1</sup>. Compound **1a** is an efficient inhibitor of the biosynthesis of peptidoglycans of bacterial cell-walls by interacting with the enzyme(s) that catalyse the transfer of the disaccharide unit from the disaccharide–(oligopeptide)–pyrophosphoryl–undecaprenol intermediate to the growing, linear peptidoglycan chain<sup>2</sup>.

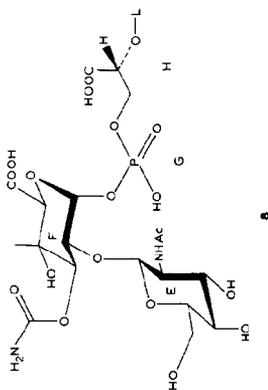
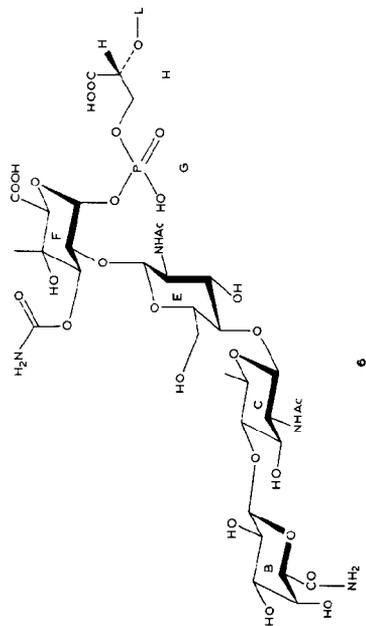
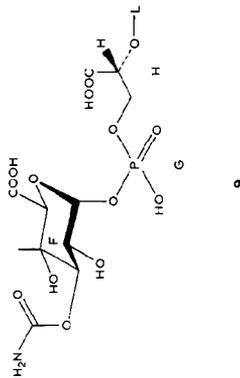
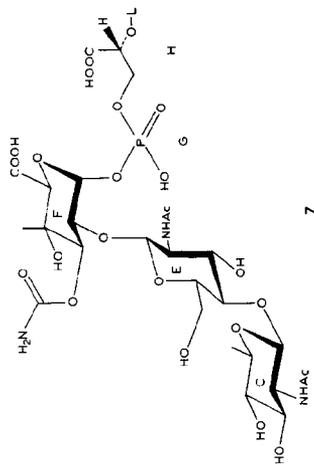
We now report on a systematic degradation of the oligosaccharide part of **1a** which was performed with the aim of defining the structural basis of the antibiotic activity of **1a**.



Transformation of **1a** to **3** by catalytic hydrogenation (to give **2**) and subsequent ozonolysis has already been described<sup>3</sup>. Compound **3** has now been obtained much more efficiently (88% yield) from **2** (7.4 mmol) by oxidation with  $K_3[Fe(CN)_6]$  (80.8 mmol) in 0.37M  $K_2CO_3$  and subsequent reversed-phase chromatography (HP 20, water–methanol gradient). We believe that an intermediate of type **1b** is involved in this reaction.

\*To whom enquiries should be addressed.





L = Lipid part (see unit I of 2)

Stepwise degradation of **3** was based on the diol-cleavage methodology<sup>4</sup>. Compound **3** (0.72 mmol) was treated for 52 h at 90° with benzaldehyde (28 mmol) and anhydrous zinc chloride (9.9 mmol) in dimethyl sulfoxide solution<sup>5</sup>, to give, after chromatographic separations (first on HP-20 with a water–methanol gradient and then on silica gel with 10:6:1 chloroform–methanol–water), **4** in 14% yield; **4** is possibly a mixture of two diastereoisomers isomeric at the acetal carbon of the benzylidene group of unit B. Only unit D of **4** contains a free diol group. Therefore, (a) cleavage of **4** (0.12 mmol) with sodium metaperiodate (0.4 mmol) in 50% acetic acid containing sodium acetate (1.9 mmol) at 40° for 5 h, (b) reversed-phase chromatography (HP 20, water–methanol gradient) and lyophilisation, (c) reaction<sup>6</sup> (Barry degradation<sup>7</sup>) with a 100-fold excess of *N,N*-dimethylhydrazine in 2-propanol–M H<sub>2</sub>SO<sub>4</sub> at pH 4 (2.5 h at 20° and 1 h at 90°), and (d) lyophilisation and chromatographic separation (silica gel, 10:6:1 chloroform–methanol–water) gave **5** in 37% overall yield.

Liberation of another diol grouping was achieved by removing the benzylidene group from **5** by hydrogenolysis (in 1:8 methanol–acetic acid, Pd/C catalyst) to give **6** quantitatively. Degradation under the conditions described above converted **6** into **7** (32%).

TABLE I

SELECTED <sup>13</sup>C-N.M.R. DATA [δ VALUES, (CD<sub>3</sub>)<sub>2</sub>SO] FOR COMPOUNDS 3–9

3	4	5	6	7	8	9	Assignment	Unit
173.1	171.6	172.5	<sup>a</sup>	173.3	173.3	<sup>a</sup>	C-1	H
171.7	171.3	171.8	171.6	171.7	172.1	172.6	C-6	F
170.6	170.6	169.6	170.7	—	—	—	C-6	B
169.7	169.7	169.2	169.3	169.4	169.4	—	NHCOCH <sub>3</sub>	C,E
156.5	156.7	156.6	156.7	156.6	156.3	158.0	OCNH <sub>2</sub>	F
103.6	104.2	—	—	—	—	—	C-1	D
102.9	103.7	102.8	103.8	—	—	—	C-1	B
101.9	102.9	102.3	102.5	102.7	102.9	—	C-1	E
101.2	102.5	101.7	101.5	101.9	—	—	C-1	C
—	101.5	101.3	—	—	—	—	C-7 <sup>b</sup>	B
—	101.0	—	—	—	—	—	C-7 <sup>b</sup>	D
93.7	94.3	93.8	94.1	94.2	94.3	94.5	C-1	F
84.4	84.8	84.6	84.4	75.4	—	—	C-4	C
80.7	80.7	80.7	80.8	80.4	70.5	—	C-4	E
71.9	70.8	70.8	59.6	59.2	60.7	—	C-6	E
70.3	80.7	80.7	70.5	—	—	—	C-4	B
69.0	80.7	—	—	—	—	—	C-4	D
55.5	55.7	55.5	55.5	55.5	55.1	—	C-2	E
55.0	54.6	54.6	54.3	54.3	—	—	C-2	C
23.1	23.4	<sup>a</sup>	23.4	23.2	23.3	—	NHCOCH <sub>3</sub>	C,E
17.3	17.3	17.4	17.4	18.0	—	—	C-6	C
16.2	16.2	16.3	16.4	16.4	16.9	16.7	CH <sub>3</sub> -4	F

<sup>a</sup> Not observable. <sup>b</sup> Acetal carbon.

The tetrasaccharide **7** could be obtained from **3** directly by removing units B and D in one step. Thus, (a) cleavage of **3** (0.7 mmol) with an excess of sodium metaperiodate (4.7 mmol) in 50% acetic acid containing sodium acetate (7.3 mmol) for 2 h at 40°, (b) reversed-phase chromatography (HP 20, water–methanol gradient) and lyophilisation, (c) treatment of the oxidation product with a 20-fold excess of *N,N*-dimethylhydrazine in 2-propanol–M H<sub>2</sub>SO<sub>4</sub> (pH 4.5) for 3 h at 85°, and (d) chromatographic separation (first on HP 20 with a water–methanol gradient and then on silica gel with 10:6:1 chloroform–methanol–water) furnished **7** in a 47% overall yield. Compound **7** could be submitted to another degradation cycle in which ammonia was used instead of *N,N*-dimethylhydrazine<sup>8</sup>. Thus, (a) treatment of **7** (0.4 mmol) with sodium metaperiodate (1.6 mmol) under the conditions described above, (b) destruction of the excess of periodate with 1.9 mmol of ethylene glycol (1 h at 20°), (c) degradation of the oxidation products with ammonia (pH 9.5, 16 h at 20°), (d) adjustment to pH 6 and chromatographic separations (HP 20, water–methanol gradient; and then silica gel, 10:6:1 chloroform–methanol–water) gave **8** (31%) and **9** (16%). Under the same conditions, **8** was degraded to **9** in 46% yield. All new compounds gave spectra that were in agreement with the assigned structures. The most characteristic and informative <sup>13</sup>C-n.m.r. signals are collected in Table I.

In the *in vitro* assay<sup>2</sup> for the transglycosylation step, moenomycin A (**1a**), **2**, **3**, and the degradation products **6**–**8** showed the same inhibitory effect, whereas **9** was 10-fold less efficient. From these results, it is concluded that only units E, F, G, H, and I of **1a** are essential for full biological (*in vitro*) activity.

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